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Determination of Secondary Nanostructure of Model Membrane from Local Fishes for Application in Pharmaceutical Field

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As it is difficult to obtain human stratum corneum for testing the pharmacological effect of the drugs, many researchers have tried to find alternative membrane for this purpose, instead of the human skin. Many animal skins e.g. pig, reptile, or fish skin seem to be suitable for this purpose. Therefore this project will provide more details to help understand more deeply the structure of these alternative skins. This project is focused on the fish skin because such skin can be easily found in Germany. Two different techniques i.e. small and wide angle X-ray scattering (SWAXS) and differential scanning calorimetry (DSC) have been applied for characterization of some local fish skins. The results showed that different skins (i.e. human stratum corneum, pig, shed snake, and fish) showed small differences in SWAXS patterns as well as in DSC thermograms. The drug paracetamol and the surfactant sodium lauryl sulphate can penetrate into the fish skins and they can affect a rearrangement of the inner structure which contains different lipids and proteins. The data are in agreement with those obtained for the shed snake skins in our previous study.

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1. Introduction

There is a considerable interest for the use of alternative model membrane for in vitro pharmaceutical applications. However, it is particularly difficult to obtain human skin for in vitro experiments and it is therefore important to have alternative membranes which mimic human skin (Stratum corneum). It is mentioned in the literature [1–3] that different membranes can be used likewise, as for example pig, reptile, or fish skin. Our previous work showed that the local snake's shed skin from Python molurus molurus was the most effective model membrane for this purpose, because it can be easily obtained from the reptile shop without any cost. However, further model membranes e.g. from fishes may also be of interest. Some researchers [2–3] have mentioned the possibility to make use of them. The advantages of the fish skins are that they can be obtained at the supermarket or fish shop. We do not need to wait for the shedding process, as it is in case of snakes. However, the fish skins should be freshly prepared and used directly without storing. In order to understand the secondary nanostructure of model membrane, the SWAXS and DSC techniques were applied.

2. Materials and methods

Different local fish skins i.e. European plaice (*Pleuronectes platessa*), Atlantic salmon (*Salmo salar*),



Fig. 1. Examples of some DSC-thermograms of untreated fish skins and those treated with a drug: (a) gilt-head bream, wet skin, no treatment, (b) gilt-head bream, dry skin, no treatment, (c) gilt-head bream skin, treated with paracetamol, (d) trout skin, scale part no treatment, (e) salmon skin treated with ibuprofen, (f) salmon skin treated with paracetamol, (g) plaice skin black side treated with paracetamol, (h) plaice skin white side treated with paracetamol.

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Atlantic cod (Gadus morhua), Trout (Salmo trutta), Gilthead bream (Sparus aurata) from Germany were used in this project. Different substances (paracetamol, ibuprofen, sodium lauryl sulphate-SLS, Tween80) were studied to understand the effect on the model skins. The DSC measurements were performed using Perkin Elmer 7 (USA). The sample of each skin of about 10–20 mg was put into an aluminum pan which afterwards was sealed and measured under nitrogen gas in the temperature range of 30–300 °C at the scanning rate of 10 °C/min. The SWAXS measurements were performed at the beamline B1, HASYLAB, DESY, Hamburg at the X-ray energy of 14 keV. Further details of methods can be found in our previous article [1].

TABLE I DSC results showing the melting peaks as mean and standard deviation values (°C) of the pure drug or of fish skins after treatment with the respective drug $(n = 1 \div 5)$.

Fish Skin	$T_{\mathrm{Mean}},$	$T_{\rm SD},$					
(+ drug)	[°C]	[°C]					
Paracetamol	170.80	0.79					
Ibuprofen	76.61	0.50					
Gilt-head bream							
wet	61.52 & 153.34						
dry	137.25	7.65					
+ Paracetamol	116.24 &159.73	1.51 & 3.76					
Trout							
wet	51.27 &157.62	1.15 &13.37					
dry	126.28	3.01					
scale	118.83	6.56					
+Paracetamol	113.57 & 170.24	0.42 & 3.86					
Atlantic cod							
wet	47.00 &135.03						
dry	122.61	0.90					
+Paracetamol	107.95 & 161.42	$0.72 \ \& 14.52$					
Atlantic salmon							
wet	49.79	0.45					
dry	58.70 & 119.80						
scale	74.09 &114.79						
+Paracetamol	107.96 & 167.05	$1.26 \ \& 2.12$					
+Ibuprofen	135.27 & 169.15	9.62 & 4.66					
European plaice							
black side wet	49.80 &147.50						
white side wet	49.33 & 150.52						
black side dry	132.13	0.35					
white side dry	130.62	0.96					
black side+Paracet.	109.18 &178.83	0.42 & 3.69					
white side+Paracet.	111.11 &169.50	$0.41 \ \& 6.35$					

3. Results and discussion

The results of DSC measurements are demonstrated in Fig. 1. The DSC-thermograms show the melting points or re-crystallization process depending on skin or drug. Surprisingly the DSC-thermogram of the fish scale showed a good sharp peak (e.g. Fig. 1d), whereas the

TABLE Π	
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SWAXS results for scattering vectors Q of different skins and effect of surfactants. HSC – Human stratum corneum, PMM – Python molurus molurus.

Type of skin	Peak 1	Peak 2	Peak 3	Peak 4
(+ substances $)$	[1/Å]	[1/Å]	$[1/\text{\AA}]$	$[1/\text{\AA}]$
HSC, dry	0.186	0.658	1.520	1.690
Plaice black side, wet	-	0.544	1.375	2.197
Plaice white side, wet	-	0.544	1.389	2.190
Atlantic cod, wet	-	0.544	1.396	2.240
Salmon, wet	0.157	0.557	1.396	2.177
PMM, dry	0.157	0.666	1.410	1.660
Pig ear, dry	0.107	0.644	1.509	-
Pig abdomen, dry	0.121	0.659	1.375	1.503
Gilt-head bream, wet	0.265^{*}	0.529	1.382	2.183
Trout, wet	-	0.551	1.389	2.190
Salmon+5%Tween80	0.558	1.389	1.839	2.240
Salmon+5% SLS	0.150, 0.565	1.403	-	2.240
Salmon+10%Tween80	0.558	1.382	-	2.205
Atlantic $cod+5\%$ Tween80	0.565	1.389	1.847	2.283
Atlantic $cod+5\%$ SLS	0.329*, 0.536	1.453	1.839	2.219
Atlantic cod+10%Tween80	0.572	1.375	1.839	2.247
Salmon + Ethanol	0.536	1.382	-	2.190
Atlantic cod+Ethanol	0.543	1.389	1.832	2.226
Plaice white side, dry	0.988^{*}	1.389	-	2.219
Plaice black side, dry	0.644	1.375	-	2.219
Salmon, dry	0.966*	1.367	1.818	2.233
Pig abdomen $+5\%$ SLS	0.164,	1.446	1.524	1.997
	0.322*, 0.487			
Pig abdomen+Ethanol	0.107	0.616	1.496	-
$Pig~abdomen{+}5\% Tween{0}{0}$	0.107	0.623	1.510	-
$5\% \mathrm{Tween80}$	0.136	-	-	-
5%SLS	1.178	-	-	-

*Not comparable with other values.

whole skins showed many peaks (e.g. Fig. 1c, 1g). Table I shows that the wet fish skin did not produce reasonable DSC thermograms because of the effect of water from the samples. Therefore the skin samples were dried in a desiccator with an absorbing substance for at least overnight period. After application of the drying process the skins showed better DSC-curves. By comparing the melting peak of the pure drug (i.e. paracetamol or ibuprofen) and the skins treated with a drug, it can be deduced that the paracetamol drug can penetrate through all studied fish skins. However, tha secondary nanostructure of some fish skins has changed, which can be monitored by the reduction or the increase of melting peaks of the paracetamol drug. The SWAXS scattering patterns (Fig. 2) showed typical peaks at different scattering vectors (Q). After comparison of the patterns of different skins i.e. human, pig, shed snake, or fish skins, small differences were detected. The effect of the tested surfactants or enhancers, i.e. SLS and Tween80, was also clearly seen. SLS can penetrate into the model fish skins but Tween80 cannot. Some examples of SWAXS patterns are shown in Fig. 2 and the Q-values are summarized in Table II.



Fig. 2. SWAXS scattering patterns of different skins and effect of different substances: (a) pig abdomen skin, dry, no treatment, (b) pig abdomen skin treated with 5% SLS, (c) Python molurus molurus skin, dry, no treatment, (d) salmon skin treated with 5% Tween80, (e) salmon skin treated with 5% SLS, (f) 5% SLS, (g) 5%Tween80, (h) human stratum corneum.

4. Conclusions

By combination of the DSC and SWAXS characterization techniques a deeper understanding of the secondary nanostructure of the model membrane from local fishes can be obtained. Though the fish skins showed small differences in comparison with the human startum corneum, pig or shed snake skins, the results showed that the fish skins may be suitable for application in pharmaceutical field with small limitations.

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